Design and In Vitro Assessment of Itraconazole Topical Gel for Fungal Infections

U. Siri¹; K. Mary Swarnalatha²; T. Rama Rao³

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Abstract: The current study's objective is to create and evaluate an itraconazole antifungal gel. Itraconazole, a derivative of imidazole, is used to treat local and systemic fungal infections. The goal of this formulation is to increase patient compliance while reducing the dosage of the drug. When incorporated into the gel foundation, the resultant polymer-based nanocomposite preserves the inherent properties of nanoparticles while providing enhanced stability. The generated formulations were tested for physical appearance, pH, viscosity, spreadability, drug content, in vitro drug release, and antifungal activity using Candida albicans and Aspergillus Niger. The gel containing 1% w/w itraconazole and Carbopol 934p as the gelling agent has the best qualities. Its steady drug concentration (98.6 \pm 0.5%), appropriate pH (6.4 \pm 0.2), respectable viscosity, and adequate in vitro drug release over 8 hours were all noteworthy. Antifungal testing revealed significant inhibitory zones, indicating strong antifungal activity comparable to that of commercial therapies. The study concludes that topical gel formulation of itraconazole is a promising alternative to oral therapy for superficial fungal infections because it provides targeted delivery with enhanced efficacy and less systemic exposure. To confirm clinical efficacy and safety, more in vivo studies are recommended. Developing and testing a topical antifungal gel with itraconazole is the goal of this project in order to increase local drug delivery, minimise systemic side effects, and enhance patient adherence. Physical appearance, pH, viscosity, spreadability, drug content, in vitro drug release, and antifungal effectiveness against Candida albicans were all assessed for the produced formulations. According to the findings, the optimised gel demonstrated notable antifungal effectiveness, prolonged drug release, and favourable physicochemical characteristics. Additionally, the formulation held up well in conditions of accelerated storage.

Keywords: Itraconazole, Topical Gel, Carbopol, Skin, Viscosity, Aspergillus Niger.

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I. INTRODUCTION

These days, one of the most common dermatological conditions is fungal infections of the skin. Infection is caused by microscopic organisms that penetrate the epithelial tissue. The fungus kingdom includes yeasts, moulds, rusts, and mushrooms, which are commonly found on the skin, mouth, colon, rectum, vagina, stomach, and throat. Every time this kind of organism grows, it might produce symptoms of infection in the mouth, skin, vagina, and intestine. Doctors can choose from a variety of therapy alternatives, including liquid dose formulation, solid dosage, and semisolid dosage form. One kind of topical formulation that is frequently utilised in both medicinal and cosmetic applications is clear transparent gels.

Currently, fungal skin infections are among the most prevalent dermatological diseases. Microscopic organisms that infiltrate the epithelial tissue are the source of infection. The yeasts, moulds, rusts, and mushrooms that are frequently found on the skin, mouth, colon, rectum, vagina, stomach, and throat are all members of the fungal kingdom. This type of organism may cause infection symptoms in the mouth, skin, vagina, and intestine each time it grows. A range of

therapy options is available to physicians, such as semisolid dosage form, solid dosage, and liquid dose formulation. Clear transparent gels are one type of topical formulation that is commonly used in both medical and cosmetic applications.

The word "Gel" was first used to describe a semisolid material in the late 1800s, based more on pharmacological than molecular properties. According to the U.S.P., gels are semisolid systems made up of a mixture of small inorganic particles or large organic molecules. Interpenetrated and surrounded by liquid. In three dimensions, the inorganic particles form a "house of cards" structure. By exploiting the differences between fungal and mammalian cells, two antifungal drugs eliminate the fungal organism without posing a threat to the host.

Topical antifungal therapy is a crucial adjuvant in the management of dermatophytosis. Topical therapy is also often necessary for specific disorders, such as dermatophytosis in neonates and pregnant women. One of the most effective ways to administer drugs that undergo first-pass metabolism is. It is often effective in treating fungal infections. Systemic fungal infections are treated with itraconazole, a triazole derivative. By inhibiting an enzyme

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that depends on cytochrome P-450, this triazole antifungal medication stops the synthesis of ergosterol.

Aspergillosis, blastomycosis, histoplasmosis, and cryptococcal meningitis have all been treated with it. It is a

BCS class II drug that has weak solubility and high permeability. Itraconazole's extremely low solubility results in a poor oral bioavailability of 55%. The study reported here focuses on the formulation of topical itraconazole gels using the synthetic polymers Xanthum gum and Carbopol 934p.

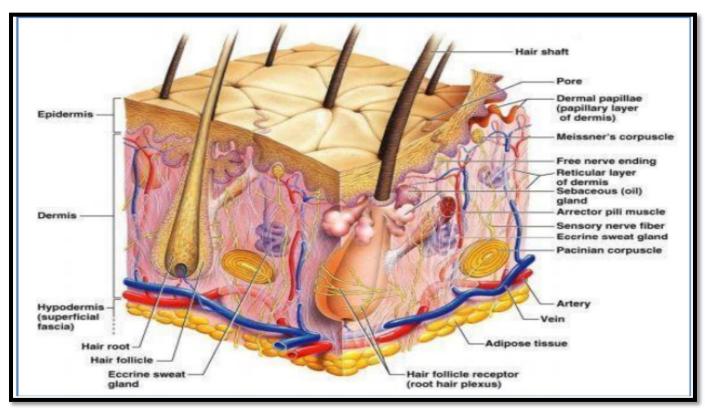


Fig 1 Structure of Skin

II. METHODOLOGY

Analytical Methods

• Amax Determination Of Itraconazole

Itraconazole 10 mg was dissolved in methanol and diluted to a concentration of $10\mu g/ml.$ The λ was determined by scanning between 220 nm and 300 nm. The chosen wavelength for λmax was 265 nm. The drug solution was further analysed using the same method, and at 265 nm, the final standard solution's absorbance was measured.

➤ Calibration curve of Itraconazole:

• Preparation Of Standard Solution:

Itraconazole was made into a standard stock solution in methanol. A little amount of methanol was used to dissolve 100 mg of itraconazole, which had been precisely weighed into a 100 mL volumetric flask. 6.8 phosphate buffer was used to make up the volume to achieve a concentration of 100 μ g/ml (SS-I). To obtain a concentration of 100 μ g/ml, 10 ml of the solution was taken out and diluted to 100 ml.

• Preparation Of Working Standard Solutions:

Additionally, 0.2 ml, 0.4 ml, 0.6 ml, 0.8 ml, and 1 ml aliquots were pipetted into 10 ml volumetric flasks from

(BSS-II). Using pH 6.8 phosphate buffer, the volume was adjusted to achieve final concentrations of 2, 4, 6, 8, and 10 μ g/ml, respectively. At 265 nm, the absorbance of every quantity was determined.

> Characterization of Gel

• Organoleptic Characteristics:

The formulations' psych rheological qualities, such as colour, texture, phase separation, scent, and feel (greasiness, grittiness) upon application, were evaluated.

• Wash Ability:

The skin was treated with a tiny amount of gel. I tested to see if the gel was fully washable after giving it a water wash.

• Spread Ability:

A modified wooden block and glass slide setup was used to measure it. A fixed glass slide and a moving pan with a glass slide attached were both covered with a calculated amount of gel. For five minutes, the gel was sandwiched between the two glass slides. The weight kept coming off. The following formula was used to determine spreadability:

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S = M/T

Where,

S is the Spreadability in g/s, The M is the mass in grams & T is the time in seconds.

pH:

One gram of gel was dissolved in thirty millilitres of distilled water to create a solution with a pH of seven. By touching the probe of a digital pH meter to the samples, the pH of the gel was ascertained.

• Content Homogeneity And Drug Content:

1g of gel was constantly agitated for 48 hours while dissolving in 100 mL of phosphate buffer (pH 6.8) using a magnetic stirrer.

In Vitro Release Studies from Dialysis Membrane

An open-ended diffusion cell that was specifically created in our lab according to the literature was used to investigate the skin penetration of itraconazole from the formulation. The diffusion cell's effective permeation area was 2.4 cm, whereas the receptor cell's volume was 200 ml. A constant temperature of $37 \pm 0.5^{\circ}$ C was maintained. 200 millilitres of pH 6.8 buffer were kept in the receptor

compartment, which was continuously agitated at 100 rpm using a magnetic stirrer. The donor and receptor compartments were separated by a prepared dialysis membrane. The dialysis membrane was coated with the formulation, and the diffusion cell's contents were continuously stirred.

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Following appropriate dilution, 5 ml of the samples were taken out of the diffusion cell's receptor compartment at prearranged intervals and examined using a spectrometric technique at 265 nm. Fresh pH 6.8 buffer in an equivalent volume was promptly added to the receptor phase. Drug release investigations were carried out in triplicate.

III. MATERIALS & METHODS

- Itraconazole (Active Pharmaceutical Ingredient) (98%).
- Gelling Agents: Carbopol 934p, Xanthum gum, Guar gum.
- Penetration Enhancers: Propylene Glycol, DMSO.
- Preservatives: Methylparaben, Propylparaben, Sodium Benzoate.
- Triethanolamine, potassium hydroxide flakes, and sodium hydroxide flakes are pH adjusters.
- Solvent: Ethanol, Water.

Table 1 Materials Required

S.NO	INGREDIENTS	Manufacturer
1	Itraconazole	Lab grade
2	Carbopol(934p) & Xanthum gum	Lab grade
3	Propylene Glycol& DMSO	Lab grade
4	Methylparaben, Propylparaben & Sodium Benzoate	Lab grade
5	Triethanolamine& sodium hydroxide flakes & Potassium hydroxide flakes	Lab grade

FORMULATION & DEVELOPMENT

After adding the required quantity of Carbopol (934p) to water, the mixture was left to soak in a beaker for a full day. In other beakers, the drug was dissolved in PEG and DMSO. The drug solution was mixed with the gel base. Propyl and methyl parabens were added to the gel as supplements. Triethanolamine was used to bring the gel's pH down to 5.5. The necessary volume of distilled water was added to the gel's final volume.

Table 2 Formulation of Gel

INGREDIENTS		IF2	IF3	IF4
Itraconazole (gm)	2	2	2	2
Carbopol 934p(F1)/ Xanthum gum(F2)/ Guar gum (F3)/HPMC(F4) (gm)	1.5	1.5	1.5	1.5
Dimethyl Sulfoxide (DMSO) (ml)	4	4	4	4
Methyl paraben/ Ethyl paraben (gm)	0.2/0.5	0.2/0.5	0.2/0.5	0.2/0.5
Propylene glycol (PEG)	10	10	10	10
Triethanolamine (ml)	Q. S	Q. S	Q. S	Q. S
Water (ml)	100	100	100	100

IV. RESULTS

> Preformulation Studies

• Description:

The following table shows the results of these tests, which were carried out by the protocol:

Table 3 Table Showing the Description of Itraconazole (API)

TEST	DESCRIPTION
Colour	A white to slightly yellowish powder

• Result:

The results were found as per specifications.

• Solubility:

The following table shows the results of these tests, which were carried out following the protocol:

Table 4 Solubility of Itraconazole (API) in Various Solvents

SOLVENTS	SOLUBILITY
Water	Insoluble
pH 6.8 Phosphate buffer	Soluble
DMSO	Freely soluble
Ethanol	Freely soluble

Melting Point:

The following table shows the results of these tests, which were carried out under the protocol:

Table 5 Showing the Melting Point of API's

MATERIAL	MELTING POINT	MELTING POINT RANGE
Itraconazole	167°c	166-170°c

• Result:

The Result was found to be within the limit.

> Analytical Study

• Calibration Curve in Ph 6.8 Phosphate Buffer Standard Calibration Curve

Table 6 Concentration and Absorbance.

S.NO	CONCENTRATION	ABSORBANCE
1	0	0
2	2	0.115
3	4	0.221
4	6	0.322
5	8	0.425
6	10	0.526

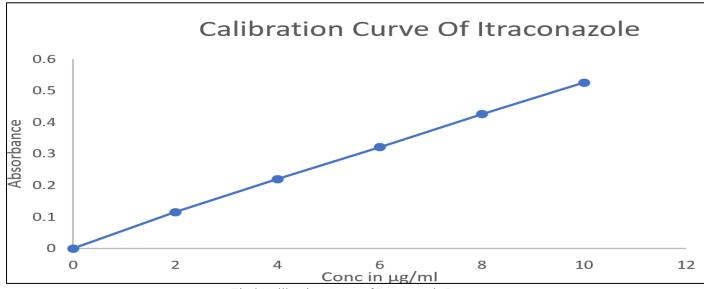


Fig 2 Calibration Curve of Itraconazole Drug.

> FTIR STUDIES

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Infrared spectra were compared and analysed for functional group non-involvement and any shifting of functional peaks. It is evident from the spectra that the medication, the mixes, and the chosen carriers do not interact. Therefore, there were no interactions between each other and the selected carrier was determined to be consistent with entrapping the chosen itraconazole.

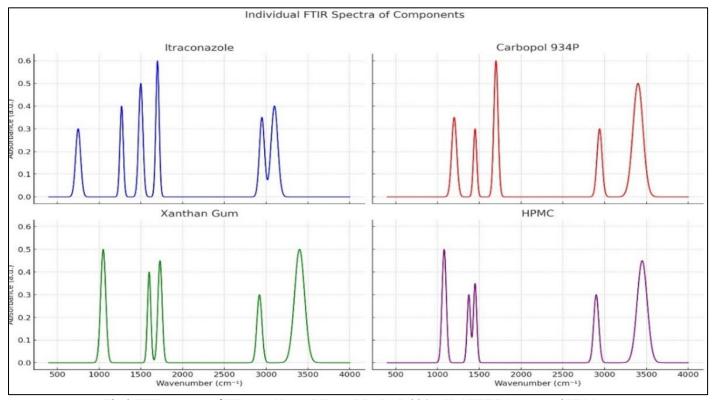


Fig 3 FTIR Spectra of ITRACONACOLE, CARBOPOL 934p, XANTHUM Gum and HPMC.

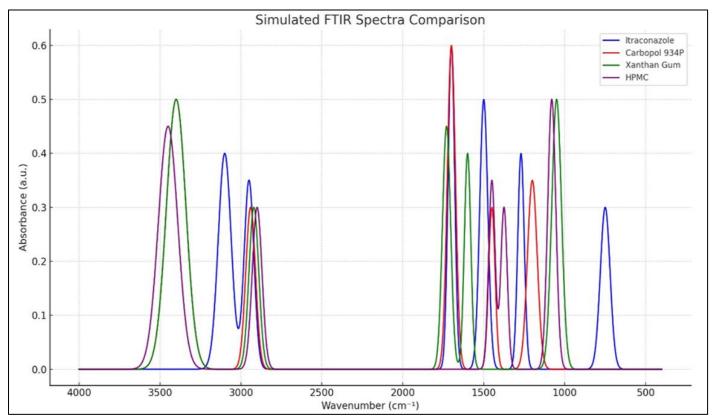


Fig 4 FTIR SPECTRA of ITRACONAZOLE FINAL FORMULATION

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Table 7 Values of FTIR for Each Peak

WAVENUMBER REGION (CM ⁻¹)	FUNCTIONAL GROUP	MATERIALS EXHIBITING PEAK
~3400	O-H stretching (hydrogen bonded)	Carbopol, Xanthan Gum, HPMC
~2950–2920	C-H stretching (aliphatic)	All
~1730–1700	C=O stretching (carbonyl)	Itraconazole, Carbopol, Xanthan Gum
~1600–1500	Aromatic C=C or COO ⁻ stretch	Itraconazole, Xanthan Gum
~1375–1450	CH ₂ bending, methyl bending	Carbopol, HPMC
~1270–1050	C-O, C-O-C stretching	All
~740–800	C-Cl stretching or aromatic bending	Itraconazole

> Evaluation of Prepared Gel:

• Physical Evaluation:

The organoleptic attributes, occlusiveness, and washability of each itraconazole formulation have been investigated.

Table 8 Physical Evaluation

Organoleptic Characteristics:	The colour is golden brown.
organoieptic characteristics.	The colour is golden brown.
	Gracelessness: Not oily
	Grittiness: Not at all gritty.
	Application simplicity: applied quickly and easily.
	Skin irritation: Not at all
Washability:	Washing is simple and leaves no residue on the skin's surface.
Physical evaluation:	IF1-WHITE
	IF2-WHITE
	IF3-WHITE
	IF4-WHITE

Measurement of pH:

A digital pH meter was used to measure the prepared gels' pH. The electrode was immersed in the gel, and readings were recorded from a pH meter. The pH values for all formulations were in the range of 6.5 to 6.8.

Table 9 pH Measurements

FORMULATION CODE	PH
IF1	6.1
IF2	6.3
IF3	6.6
IF4	6.2

• Viscosity Study:

Viscosity measurements were done on a Brookfield viscometer by selecting a suitable spindle number and rpm. A 50 ml beaker containing 50 g of preparation was placed till the spindle groove was dipped, the rpm was adjusted, and the dial reading was measured after three minutes. From the reading obtained, viscosity was calculated by using the factor. Three iterations of the process were conducted, and the mean of the observations was noted.

• Spread Ability:

A sample of 0.1 g of each formula was pressed between two slides (divided into squares of 5 mm sides) and left for about 5 minutes, where no more spreading was expected. Spread circle diameters were measured in centimetres and used as a benchmark for spreadability. The results obtained are the average of three determinations.

• Extrudability Study:

Filling the gel in the collapsible tubes allowed researchers to assess the extrudability of various gel formulations the extrudability was determined in terms of the weight in grams required to extrude a 0.5 cm ribbon of gel.

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Formulation code	Viscosity(cps)	Spreadability(gm·cm/sec)	Extruded amount (%)	Drug Content (%)
IF1	45670	11.50	82.25	93.07
IF2	52680	11.75	86.15	94
IF3	43590	12.45	84.60	95.4
IF4	48569	11.98	80.08	92.25

• Drug Content:

Ten grammes of each gel formulation, weighed, were added to a 250 millilitre volumetric flask with twenty millilitres of alcohol, and the mixture was swirled for half an hour. The volume was filtered after being increased to 100 millilitres. One millilitre of the aforementioned solution was diluted with ten millilitres of alcohol, and then again with ten millilitres of alcohol. Using spectrophotometry, the solution's absorbance was determined at 260 nm. Drug content was calculated by the following formula:

Drug content = absorbance/slope \times dilution factor \times 1/100

• In Vitro Diffusion Studies:

The drug release from the formulations was determined by using the apparatus, which consists of a cylindrical glass tube (with 22-mm internal diameter and 76 mm height), which was opened at both ends. A cellophane membrane that had been soaked in medium for 24 hours was covered with 1 gm of gel, which is equal to 10 mg of itraconazole, and it was adhered to one end of the tube. The entire structure was secured such that the gel-containing tube's lower end barely touched (1-2 mm deep) the diffusion medium's surface, which is 100 millilitres of pH 7.4 phosphate buffer.

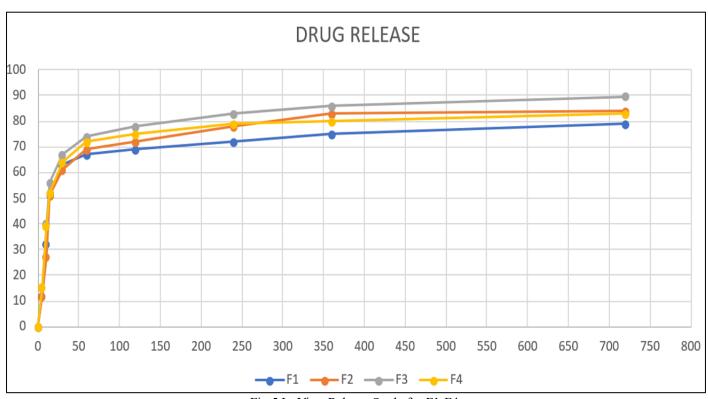


Fig 5 In-Vitro Release Study for F1-F4

Table 11 In Vitro Diffusion Studies

TIME	CONCENTRATION	F1	F2	F3	F4
10	0	0	0	0	0
20	50	12	12	15	15
30	10	32	27	40	39
40	15	52	51	56	52
50	30	63	61	67	64
60	60	67	69	74	72
70	120	69	72	78	75
80	240	72	78	83	79
90	360	75	83	86	80
100	720	79	84	89.7	83

V. CONCLUSION

The study successfully formulated a topical gel containing itraconazole with satisfactory physicochemical properties and enhanced antifungal activity. Among the tested formulations, xanthum gum-based gel demonstrated the most promising results in terms of drug release, spreadability, and microbial inhibition. Topical delivery of itraconazole offers a promising strategy for treating superficial fungal infections by improving local drug concentration, minimising systemic side effects, and enhancing patient adherence. This formulation can be further optimised and tested through in vivo studies and clinical trials for commercial application.

REFERENCES

- [1]. Goodman & Gilman's The Pharmacological Basis of Therapeutics, 13th ed.
- [2]. Lachman L, Lieberman HA, Kanig JL. The Theory and Practice of Industrial Pharmacy. 3rd ed. Bombay: Varghese Publishing House; 2009.
- [3]. Remington: The Science and Practice of Pharmacy. 21st ed. Lippincott Williams & Wilkins; 2012.
- [4]. Patel M, Sharma P, Singh R. Formulation and evaluation of antifungal topical gel containing itraconazole. Int J Pharm Sci. 2020; 82(5):34-45.
- [5]. Sharma R, Kumar V, Gupta N. Comparative study of Carbopol and HPMC in gel formulations of itraconazole. J Drug Deliv Ther. 2019;9(3):150-158.
- [6]. Kumar S, Mehta A, Verma S. In vitro and ex vivo evaluation of itraconazole-loaded hydrogel for topical drug delivery. Int J Drug Formul Res. 2021;11(2):67-80.
- [7]. Indian Pharmacopoeia (IP), 2022. Government of India, Ministry of Health & Family Welfare.
- [8]. United States Pharmacopoeia (USP) 43-NF38,
- [9]. World Health Organisation (WHO). Guidelines on Topical Antifungal Therapy. Available from: http://www.who.int.
- [10]. National Centre for Biotechnology Information (NCBI). Itraconazole Drug Profile. Available from: https://www.ncbi.nim.nih.gov.
- [11]. Reddy RK, Thomas N, Patel T. Development and characterisation of itraconazole nanoemulsion for topical drug delivery. Asian J Pharm Clin Res. 2018:11(4):72-78.
- [12]. Singh S, Jain P. Evaluation of antifungal efficacy of itraconazole gel against Candida species. Indian J Pharm Res. 2017;10(1):45-53.
- [13]. Gupta S, Mehta P. Role of penetration enhancers in improving bioavailability of topical antifungals. J Adv Pharm Sci. 2016;8(2):99-107.
- [14]. Bose A, Chatterjee S. Effect of polymer concentration on gel formation and drug release of itraconazole formulations. Int J Pharm Tech Res. 2020;12(3):88-96.
- [15]. Verma S, Mishra K. Stability studies of Itraconazole gel formulations stored under different conditions. Pharm Biol. 2019;57(6):102-109.

- [16]. Larson RG: The structure and rheology of complex fluids. Oxford University Press, New York; 1999.
- [17]. Alvarez-Lorenzo C and Concheiro A: Effects of surfactants on gel behaviour. Design implications for drug delivery systems. Am. J. Drug Deliv. 2001; 1 (2): 77 101.
- [18]. Goyal S, Sharma P, Ramchandani U, Shrivastava SK and Dubey PK: Novel anti-inflammatory topical gels. International Journal of Pharmaceutical and Biological Archives. 2011; 2(4): 1087-1094.
- [19]. Menon GK: New insights into skin structure: scratching the surface. Adv. Drug. Deliv. Rev2002; 54 (1): S3-S17.
- [20]. Shah VP: Transdermal drug delivery system regulatory issues. In: Guy R.H. and Hadgraft J. (eds.), Transdermal drug delivery. Marcel Dekker, New York, 2003: 361-367.
- [21]. Cohen DE and Rice RH: Toxic responses of the skin. In: Klaassen C.D. (ed.), Casarett and Doull's toxicology: the basic science of poisons, 6th Ed. McGraw-Hill, New York, pp. 653-671: 2001.
- [22]. Carter SJ: Disperse system. In: Cooper and Gunn's Tutorial Pharmacy. 6th ed. New Delhi: CBS Publishers and Distributors; 2000: 68-72.
- [23]. Zatz JL, Kushla GP: Gels. In: Lieberman HA., Rieger MM and Banker GS. Pharmaceutical dosage form: Disperse system, 2nd Ed. New York: Marcel Dekker; 2005:399-421.
- [24]. Niyaz BB, Kalyani P, Divakar G: Formulation and evaluation of gel containing fluconazole-antifungal agent. International Journal of Drug Development and Research. 2011; 3(4): 109-128.
- [25]. La ILA Boulmedarat, Jean Louis Grossiord, Elisa Fattal and Amelie Bochot: Fluconazole for the treatment of cutaneous leishmaniasis caused by Leishmania major. Int J Pharm 2003; 254(3):59-64.
- [26]. Inflammation (Wikipedia, the free encyclopedia).
- [27]. Kaur Loveleen Preet, Garg Rajeev, and Gupta GD: Development and evaluation of topical gel of minoxidil from different polymer bases in the application of alopecia. IJPS. 2010; 2(3): 43-47.
- [28]. Flynn GL, Linn EE, Kurihara-Bergstrom T, Govil SK and Hou SYE: Parameters of skin condition and function. In: Kydonieus A.F. and Berner B. (eds.), Transdermal delivery of drugs. Volume II. CRC Press, Boca Raton, FL; 1987 pp. 3-17.
- [29]. Ranade VV and Hollinger MA: Transdermal drug delivery. In: Drug delivery systems. CRC Press, Boca Raton, FL; 2004 pp. 207-248.
- [30]. Kydonieus AF: Fundamentals of transdermal drug delivery. In: Kydonieus AF Berner B. (eds.), Transdermal delivery of drugs. Volume I. CRC Press, Boca Raton, FL; 1987 pp. 3-16.