

Phytochemical Analysis, Antioxidant Activity, and Antimicrobial Evaluation of Kinnow Fruit Peel Extract

Nivedita S M¹; Dr. Neelakanth M Jeedi^{2*}

^{1,2}Department of Pharmacology, KLE College of Pharmacy, Hubballi - 580031 (A Constituent Unit of KLE Academy of Higher Education and Research, Belagavi, Karnataka), India

Corresponding Author: Dr. Neelakanth M Jeedi^{2*}

Publication Date: 2025/07/09

Abstract: This study investigates the bioactive constituents, antioxidant efficacy, and antimicrobial activities of kinnow fruit peel extract. Qualitative phytochemical analysis confirmed the presence of flavonoids, phenolics, tannins, and alkaloids. The extract's antioxidant capacity, assessed using DPPH assay, exhibited notable radical scavenging activity. Antimicrobial properties were tested against *E. coli* and *S. aureus* using agar well diffusion technique, showing significant zones of inhibition. These results indicate the potential of kinnow peel as a natural therapeutic agent and sustainable by-product for the development of nutraceuticals and natural antimicrobial products.

Keywords: *Phytochemicals Analysis, Antioxidant Activity, Antimicrobial Evaluation, Kinnow Fruit Peel Extract, Citrus Bioactives, Natural Therapeutics.*

How to Cite: Nivedita S M; Dr. Neelakanth M Jeedi (2025) Phytochemical Analysis, Antioxidant Activity, and Antimicrobial Evaluation of Kinnow Fruit Peel. *International Journal of Innovative Science and Research Technology*, 10(7), 140-144, <https://doi.org/10.38124/ijisrt/25jul227>

I. INTRODUCTION

Plants have long been recognized for their medicinal properties and are often considered safer than synthetic alternatives due to fewer side effects. The rising global interest in natural remedies has brought attention to citrus fruits, especially kinnow (*Citrus reticulata* × *Citrus sinensis*), known for its high vitamin C content and rich phytochemical profile. Despite its nutritional benefits, kinnow peel, often discarded as waste, contains numerous bioactive compounds. This study explores the potential of kinnow fruit peel extract for antioxidant and antimicrobial applications, aligning with global trends towards sustainable and natural therapeutics.

II. MATERIALS AND METHODS

The study employed standardized protocols for sample preparation and phytochemical screening. Kinnow fruit peels were collected, washed, shade-dried, and powdered. Solvent extraction was performed using ethanol, methanol, acetone, and distilled water. Soxhlet extraction and maceration techniques were used. Extracts were filtered and stored under refrigeration until use. Phytochemical screening involved qualitative tests for alkaloids (Mayer's and Wagner's),

flavonoids (NaOH test), phenolics (Ferric Chloride), tannins, saponins, and carbohydrates (Benedict's and Fehling's).

Antioxidant activity was determined using the DPPH assay. A 1 mM DPPH solution in methanol was prepared, and various concentrations (20–100 µg/mL) of the extract were mixed with the solution. Absorbance was measured at 515 nm after 15 minutes of incubation in the dark. IC₅₀ values were calculated. Antimicrobial activity was assessed using the agar well diffusion method. Bacterial strains *Escherichia coli* and *Staphylococcus aureus* were tested against the extract. Wells were loaded with 100, 50, and 25 mg/mL concentrations of extract. Zones of inhibition were measured after 24-hour incubation at 37°C.

III. RESULTS

Phytochemical screening confirmed the presence of flavonoids, tannins, alkaloids, phenolics, and saponins in the kinnow peel extract, indicating its rich phytochemical profile. The antioxidant activity revealed a dose-dependent increase in radical scavenging, with IC₅₀ value of 64.47 µg/mL compared to 58.66 µg/mL for standard vitamin C, demonstrating comparable efficacy.

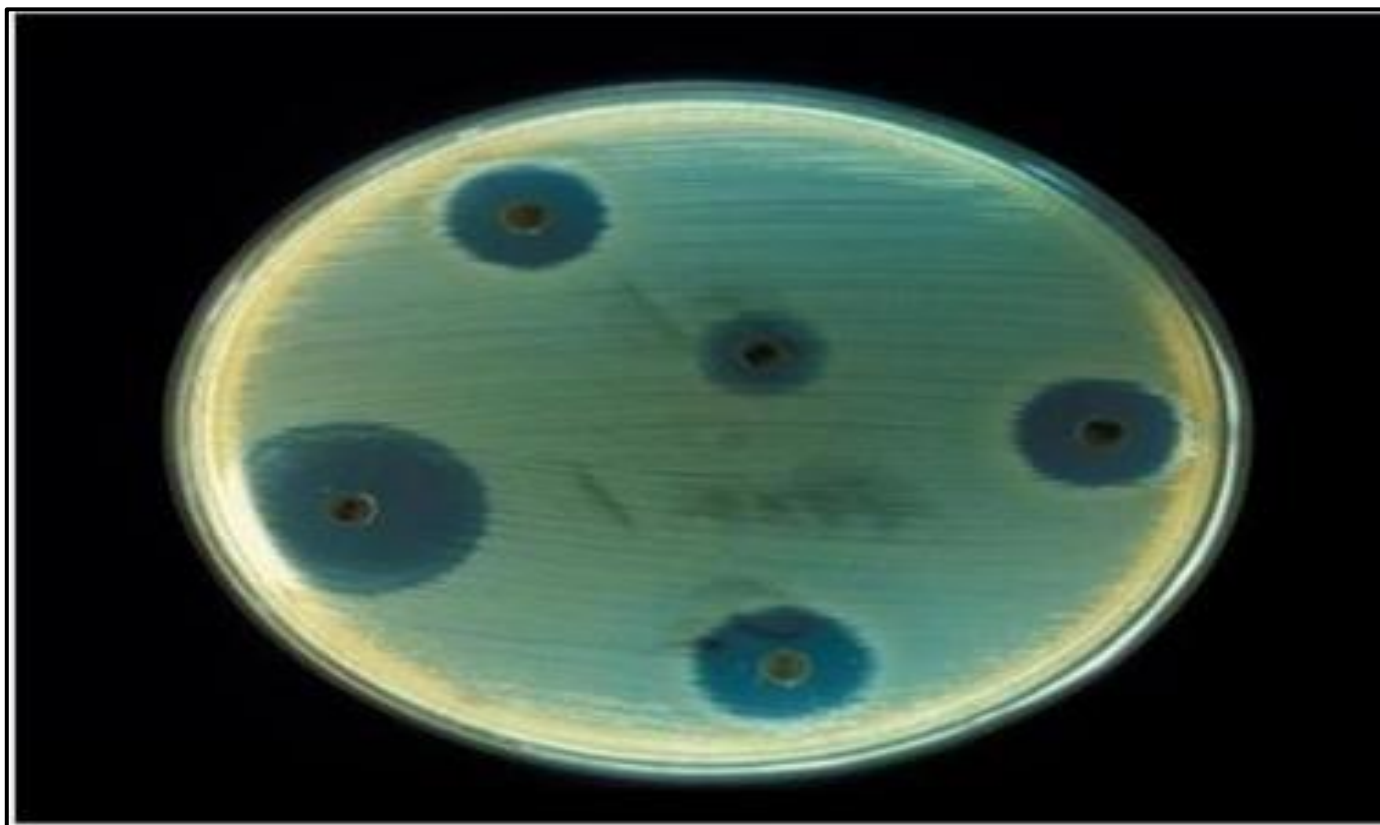


Fig 1 Zone of Inhibition – *E. coli*

This figure illustrates the antibacterial effect of kinnow peel extract on *Escherichia coli* using the agar well diffusion method. A clear zone around the well indicates microbial

growth inhibition. The highest activity was observed at 100 $\mu\text{g/mL}$ with a zone diameter of 9 mm, demonstrating the extract's potential efficacy against Gram-negative bacteria.

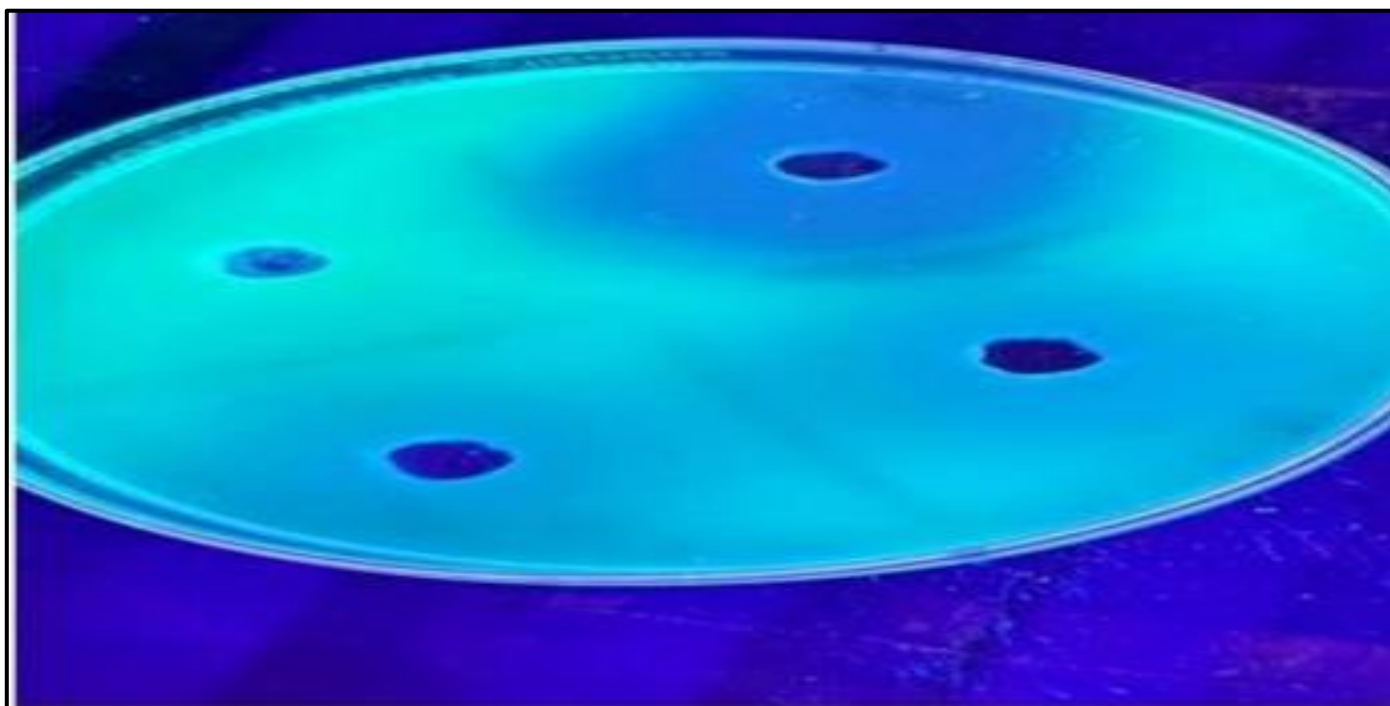


Fig 2 Zone of Inhibition – *S. aureus*

This figure shows the inhibition zone produced by the extract against *Staphylococcus aureus*. A zone diameter of 8 mm was recorded at 100 $\mu\text{g/mL}$ concentration, confirming

moderate antibacterial activity against Gram-positive bacteria. The results support the broad-spectrum antimicrobial potential of the kinnow peel extract.

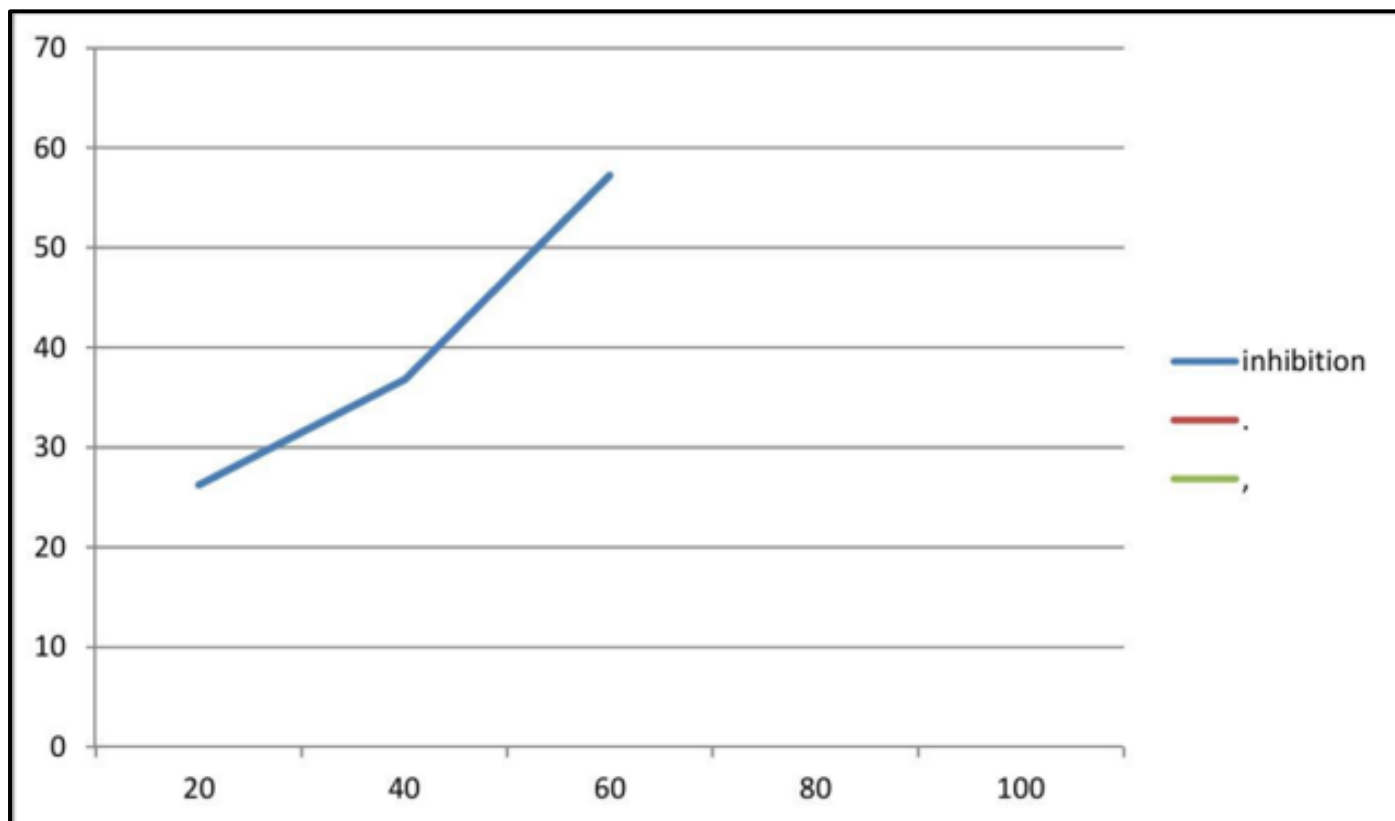


Fig 3 Free Radical Scavenging Activity (Vitamin C vs Extract)

Table 1 Comparison of Free Radical Scavenging activity between Kinnow Peel Extract and Vitamin C.

Concentration (µg/mL)	Extract % Inhibition	Vitamin C % Inhibition
20	26.25	~30
40	36.79	~42
60	57.25	~55

This figure illustrates the free radical scavenging activity of kinnow peel extract compared with standard Vitamin C (ascorbic acid), as measured by the DPPH assay. Both substances show a dose-dependent increase in antioxidant activity. The extract reached a maximum inhibition of 57.25% at 60 µg/mL, while Vitamin C achieved slightly higher inhibition (approximately 60–65%) at the same

concentration. The IC₅₀ value of the kinnow extract (64.47 µg/mL) was close to that of Vitamin C (58.66 µg/mL), indicating strong antioxidant potential. These findings suggest that kinnow peel extract can serve as a natural and effective antioxidant agent, supporting its use in functional foods and herbal formulations.

Table 2 Phytochemical Screening

S. No	Phytochemical Test	Reagent Used	Result
1	Alkaloids	Mayer's reagent	Positive
2	Phenol	Ferric chloride	Positive
3	Terpenoids	Chloroform	Positive
4	Reducing Sugars	Fehling's A & B	Positive

This table presents the qualitative results of the phytochemical analysis conducted on kinnow peel extract. Standard tests confirmed the presence of key secondary metabolites such as alkaloids, phenols, terpenoids, and

reducing sugars. These compounds are known to contribute to the antioxidant and antimicrobial properties of plant-based extracts, suggesting that kinnow peel holds substantial therapeutic potential.

Table 3 IC₅₀ values for Antioxidant activity of Kinnow Peel Extract and Standard Vitamin C

Sample	IC ₅₀ Value (mg/mL)
Vitamin C (Standard)	58.66
Kinnow Extract	64.47

This table compares the IC₅₀ values of kinnow peel extract and standard vitamin C, indicating the concentration required to scavenge 50% of DPPH free radicals. The extract showed an IC₅₀ value of 64.47 µg/mL, slightly higher than

vitamin C at 58.66 µg/mL. While less potent than the standard, the extract demonstrates significant antioxidant potential and could serve as a natural alternative to synthetic antioxidants.

Table 4 Antibacterial Inhibition Zones

Bacteria	Extract Conc. (µg/mL)	Zone of Inhibition (mm)
<i>E. coli</i>	100	9
<i>S. aureus</i>	100	8
<i>E. coli</i>	50	Moderate
<i>S. aureus</i>	50	Moderate

This table outlines the inhibitory effects of kinnow peel extract against two pathogenic bacteria—*Escherichia coli* and *Staphylococcus aureus*—at varying concentrations. A clear dose-dependent antibacterial response was observed, with the highest zone of inhibition recorded at 100 µg/mL (9 mm for *E. coli* and 8 mm for *S. aureus*). These findings validate the antimicrobial potential of kinnow peel and support its future application in natural antibacterial formulations.

IV. DISCUSSION

The antioxidant and antimicrobial efficacy of kinnow peel extract can be attributed to its rich phytochemical composition, particularly the presence of phenolic compounds and flavonoids. These compounds exhibit strong redox properties, allowing them to act as reducing agents, hydrogen donors, and singlet oxygen quenchers. Their roles in disease prevention, particularly in oxidative stress-related conditions such as cancer, cardiovascular diseases, and neurodegenerative disorders, have been well-documented in previous studies.

In the context of antimicrobial activity, the observed zones of inhibition reflect the efficacy of the extract against both Gram-positive and Gram-negative bacteria. This suggests that the extract disrupts microbial cell walls or interferes with metabolic pathways critical for bacterial survival. These findings are consistent with existing literature highlighting the antimicrobial potential of citrus-derived bioactive. Notably, the use of kinnow peel extract aligns with sustainable development goals by repurposing agricultural waste into value-added products.

While the in vitro results are promising, further studies are necessary to isolate specific active constituents and to validate these findings through in vivo models. Clinical studies will be critical in determining safety, bioavailability, and efficacy in therapeutic contexts.

V. CONCLUSION

Kinnow fruit peel, a typically discarded by-product, has shown significant pharmacological potential through its rich phytochemical content and biological activities. The study highlights its antioxidant and antimicrobial efficacy, reinforcing its potential in pharmaceutical, nutraceutical, and food industries. Further research should focus on isolation and in vivo testing of active constituents to validate therapeutic applications.

REFERENCES

- [1]. Yaqoob M., Aggarwal P., Babbar N. (2022). Extraction and screening of kinnow (*Citrus reticulata* L.) peel phytochemicals. *Biomass Conversion and Biorefinery*.
- [2]. Baswal A. K., et al. (2021). Edible coatings maintain phytochemicals in 'Kinnow' mandarin. *Australian Journal of Crop Science*, 15(10), 1332–1338.
- [3]. Purewal S. S., et al. (2022). Valorization of Kinnow peel bioactives. *Journal of Food Measurement & Characterization*, 17(1), 787–799.
- [4]. Gnanasaraswathi M., et al. (2014). Antioxidant behavior of citrus peels. *J Chem Pharmaceut Sci*, 2:139–144.
- [5]. Nimbal S. K., et al. (2024). Isolation and Identification of Cellulose producing Bacteria from Rotten Fruits. *YMER*, 23(09), 545–552.
- [6]. Rafiq, S., Singh, B., & Gat, Y. (2019). Effect of different drying techniques on chemical composition, color and antioxidant properties of kinnow (*Citrus reticulata*) peel. *Journal of Food Science and Technology*, 56(5), 2458–2466.
- [7]. Pulparambil, A., Rasane, P., Singh, J., Kaur, S., Bakshi, M., Mahato, D. K., Kaur, J., Gunjal, M., & Bhadariya, V. (2024). Bioactive Compounds from Kinnow Processing Waste and their Associated Benefits: A Review. *Recent Advances in Food, Nutrition & Agriculture*, 15(2), 103–114.
- [8]. Safdar, M. N., Kausar, T., Jabbar, S., Mumtaz, A., Ahad, K., & Saddozai, A. A. (2017). Extraction and quantification of polyphenols from kinnow (*Citrus reticulata* L.) peel using ultrasound and maceration techniques. *Journal of Food and Drug Analysis*, 25(3), 488–500.
- [9]. Saini, M. K., Neena Capalash, Varghese, E., Kaur, C., & Singh, S. P. (2022). A Targeted Metabolomics Approach to Study Secondary Metabolites and Antioxidant Activity in "Kinnow Mandarin" during Advanced Fruit Maturity. *Foods*, 11(10), 1410–1410.
- [10]. Abd-Elal H A and Halaweish F T (2010) Food preservative activity of phenolic compounds in orange peel extracts (*Citrus sinensis* L.). *Lucrari stiintifice Seria Zootehnie* 53: 233-40.
- [11]. Ahmad M M, Salim-ur-Rehman Z, Iqbal F M A and Sultan J I (2006) Genetic variability to essential oil composition in four citrus species. *Pak J Bot* 38(2): 319- 24.

- [12]. Alia M, Horcajo C, Bravo L and Goya L (2003) Effects of grape antioxidant dietary fiber on the total antioxidant capacity and the activity of liver antioxidant enzymes in rats. *Nutr Res* 23: 1251-67.
- [13]. Amarowicz R, Troszynska A, Barylko-Pikielna N and Shahidi F (2004) Extracts of polyphenolics from legume seeds- Correlation between their total antioxidant activity, total phenolics content, tannins content and astringency. *J Food Lipid* 11: 278-86.
- [14]. Amic D, Davidovic-Amic D, Beslo D and Trinajstić N (2003) Structure related scavenging activity relationship of flavonoids. *Croat Chem Acta* 76: 55-61.
- [15]. Anagnostopoulou M A, Kefalas P, Papageorgiou V P, Assimopoulou A N and Boskou D (2006) Radical scavenging activity of various extracts and fractions of sweet orange peel (*Citrus sinensis*). *Food Chem* 94: 19-25.
- [16]. Araujo C R R, Silva T M, Lopes M, Villela P, Alcantara A F C and Dessimoni-Pinto N A V (2013) Total antioxidant capacity, total phenolic content and mineral elements in the fruit peel of *Myrciaria cauliflora*. *Braz J Food Technol* 16(4): 301-09. Arts I C W, Vande-Putte B and Hollman P C H (2000) Catechin contents of foods commonly consumed in the Netherlands. *J Agric Food Chem* 48: 1752-57.
- [17]. Chou C F and Huang Y L (2003) Comparison of the chemical composition and physicochemical properties of different fibers prepared from the peel of *Citrus sinensis* L. *J Agric Food Chem* 51: 2615-18.
- [18]. Cvetnić Z and Vladimir-Knezević S (2004) Antimicrobial activity of grapefruit seed and pulp ethanolic extract. *Acta Pharm* 54: 243-50.
- [19]. D'Archivio M, Filesi C, Di B R, Gargiulo R, Giovannini C and Masella R (2007)
- [20]. Polyphenols, dietary sources and bioavailability. *Ann Ist super sanità* 43(4): 348-61.
- [21]. Dahmoune F, Boulekbache L, Moussi K, Aoun O, Spingo G and Madani K (2013) Valorization of citrus lemon residues for the recovery of antioxidants: Evaluation and optimization of microwave and ultrasound application to solvent extraction. *Ind crop prod* 50: 77-87.
- [22]. Ghasemi K, Ghasemi Y and Ebrahimzadeh M A (2009) Antioxidant activity, phenol and flavonoid contents of 13 citrus species peels and tissues. *Pak J Pharm Sci* 22(3): 277-81.
- [23]. Giulia D C, Nicola M, Angelo A and Francesco C (1999) Flavonoids: Old and new aspects of a class of natural therapeutic drugs. *Life Sci* 65: 337-53.
- [24]. Gnanasaraswathi M, Lakshmi Prabha S, Rajadurai J R P, Abhinayashree M, Fathima B M, Aarthi L V and Kamatchi S (2014) Potent anti-oxidant behavior of citrus fruit peels and their bacterial activity against multi drug resistant organism *Pseudomonas aeruginosa*. *J Chem Pharmaceut Sci* 2: 139-44.